50 μM CaCl₂ stimulated aggregation. A higher concentration of CaCl₂ seemed to be inhibitory.

Specificity of calcium was tested. Mg2+-EDTA pulses and sustained MgCl₂ application had no effect. When EGTA was used instead of EDTA, it induced csA similarly. This study shows that Ca²⁺-EDTA pulses stimulated the

formation of EDTA resistant cell contacts as well as cAMP

pulses did. As in the case of cAMP, pulse application of calcium could induce cell contacts in a shorter period than sustained application. It has been reported that 3 pulses of CaCl₂ induced spontaneous oscillation of light scattering in cell suspension. The induction of cell contact by Ca2+-EDTA pulses may be due to triggered cellular oscillations. Calmodulin⁹ may take part in this process.

- 1 Present address: Department of Anatomy, Dokkyo University School of Medicine, Mibu, Tochigi (Japan).
- C. Klein and M. Darmon, Nature 268, 76 (1977).
- G. Gerisch, H. Fromm, A. Huesgen and U. Wick, Nature 225, 547 (1975).
- U. Wick, K. Malchow and G. Gerisch, Cell Biol. int. Rev. 2, 71
- J. W. Mason, J. Rasmussen and F. Dibella, Exp. Cell Res. 67, 156 (1971).
- 6 M.H. Juliani and C. Klein, Biochim biophys. Acta 497, 369
- C. Klein and P. Brachet, Nature 254, 432 (1975).
- G. Gerisch, D. Malchow, A. Huesgen, V. Nanjundiah, W. Roos and U. Wick, in: Developmental biology, ICN UCLA symposia on molecular and cellular biology, vol. 2, p. 76. Eds D. McMahon and C.F. Fox. W.A. Benjamin Inc., 1975.
- M. Clarke, W.L. Bazari and S.C. Kayman, J. Bact. 141, 397

Antifeedant activity of precocenes and analogs on Rhodnius prolixus¹

P.D. Azambuja, W.S. Bowers², J.M.C. Ribeiro and E.S. Garcia

Department of Physiology, Universidade Federal Fluminense, C.P.183, Niteroi, R.J., 24000 (Brazil), and Department of Entomology, New York State Agricultural Experiment Station, Cornell University, Geneva (New York 14456, USA), 27 April 1982

Summary. Precocene and analogs added to the meal of 4th instar larvae of Rhodnius prolixus were tested as antifeedants. While precocene II had a strong antifeedant effect (ED₅₀=48 μ g/ml), the other compounds showed no drastic inhibition of feeding (ED₅₀>140 µg/ml). ATP, a phagostimulant, did not reverse the antifeedant action of precocene II. The mechanism of feeding inhibition is discussed.

Precocenes are plant derived substances that induce precocious metamorphosis in immature hemipterans and prevent ovarian development in some adult insects³⁻⁶. Precocenes are selectively cytotoxic to the secretory cells of the corpus allatum of the insect, thereby eliminating the production of juvenile hormone⁷⁻¹². However, Slama¹³ concluded that premature metamorphosis and ecdysis prevention induced by precocene in certain insects were due to its antifeedant action, as prothetelic insects always appeared among the larvae with the most disturbed growth, close to lethality. In fact, Slama found that precocene profoundly inhibited feeding and growth as well as oviposition in Pyrrhocoris and Dysdercus. He concluded that the latter effect was identical to that obtained with females treated by the antifeedants myristicin and elemicin. In contrast, although Bowers and Ferugia, and Bowers and Aldrich^{14,15} had noted a disinclination by adult female milkweed bugs to feed following treatment with just subtoxic dosages of precocene II, at effective sterilizing concentrations substantial feeding and weight gain occurred. In similar experiments with the milkweed bug it was demonstrated that the main difference in weight between normal and precocene-treated insects was due to the inhibition of ovarian development in the latter¹⁶. These results emphasize that the antifeedant and anti-hormonal actions of precocene are distinct and especially that the antihormonal activities are not the result of a reduction or elimination of food intake. Considering antifeedants as substances which when tasted or initially consumed can result in cessation of feeding either temporarily or permanently depending on potency, we reported recently that precocene II, added to the diet, acted as a feeding deterrent for Rhodnius prolixus, reducing the amount of blood consumed by this insect¹⁷. We now describe the inhibition of feeding induced by precocene and its analogs in the bloodsucking bug Rhodnius prolixus. Materials and methods. Precocene II, ethoxy precocene (i.e.,

7-ethoxy-6-methoxy-2.2-dimethyl chromene¹⁸) and the chromane of precocene II were synthesized by one of us (W.S.B.); precocene I was purchased from Sigma Chemical Co. All other reagents were of analytical grade. Following ecdysis, the 4th-instar larvae of R. prolixus were starved for 20-30 days and then fed on blood or artificial food containing precocene through a special feeding apparatus¹⁹ Precocenes dissolved in ethanol were added to the citrated blood or artificial diet (1 µl of Tween 80, 20 µl of ethanol plus sample, 10 ml of 30 µM ATP in 0.15 M NaCl) at doses ranging from 10 to 100 µg/ml. Control experiments showed

Table 1. Inhibition of feeding of Rhodnius prolixus by the precocenes and analogs

Substances	Effective dose (ED ₅₀) (μg/ml)	
Chromane of precocene II*	> 300	
Precocene I*	260	
Precocene II*	48	
Precocene II**	43	
Ethoxy precocene*	140	

^{*} Drugs diluted in blood; ** drugs diluted in artificial diet.

Table 2. Effect of ATP on the inhibition of feeding induced by precocene II in Rhodnius prolixus

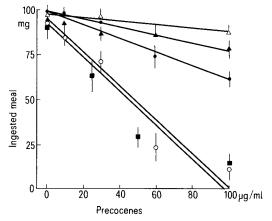
ATP (μm)	Precocene II (µg/ml)*	Meal ingested (mg)
30	_	94.5 ± 5.5
30	50	29.0 ± 6.4
300	50	36.5 ± 7.0
3000	50	34.8 ± 5.6

^{*} Artificial meal.

that ethanol at this concentration did not disturb Rhodnius feeding. The insects were allowed to feed for 30 min. The intake of blood was determined by the difference in body weight measured before and after feeding. Tests were performed on groups containing 25-30 insects. The effective antifeedant concentrations (ED₅₀) were determined by computing the linear regression, method of least squares²⁰, of the ingested meal against the dosage of precocenes.

esults and discussion. The effects of precocene I, II and derivatives on feeding are illustrated in the figure and table 1. As can be seen only precocene II showed a strong antifeedant effect. The other precocenes, as well as the chromane of precocene II, had no drastic antifeedant action at doses up to 100 µg/ml of blood. Table 1 shows that the effective antifeedant doses (ED₅₀) of chromane, precocene I and ethoxy precocene were (>140 μ g/ml) much higher than that of precocene II (48 μ g/ml of blood). The figure also shows that the effect of precocene II on the inhibition of feeding was independent of the meal quality; using artificial diet we observed the same ED₅₀ as noted for the blood meal (43 µg/ml).

Besides the results illustrated we could also observe that all precocene treatments revealed similar delayed toxicity at doses higher than 30 µg/ml of blood, while the chromane of precocene II had no toxic effect. Since it has been shown that R. prolixus is stimulated to feed by ATP²¹, we wondered what effect ATP might have on the feeding inhibition induced by precocene II. Table 2 demonstrates that the reduced meal size was an effect of precocene II and was not reversed by addition of ATP to the meal. Our data clearly indicate that change in the chemical structure of precocene II results in great loss of antifeedant activity. The 6,7dimethoxy group in the 2,2-dimethylchromene (precocene II) seems to be essential for the antifeedant property of precocene. If one of these substituents is replaced by 7ethoxy (ethoxy precocene) or H- (precocene I) the antifeedant activity is vastly decreased. Although chemical inhibition of feeding has been studied in detail for a few phytophagous insect species, the mechanism(s) of action could function by blocking the input from receptors normally responding to phagostimulants or by stimulating specific 'deterrent' cells²². It is known that the former mechanism may be reversed by increasing the amount of phagostimulants²³. In our case it was not possible to reverse the antifeedant action of precocene by increasing the ATP



Inhibition of feeding by the precocenes and derivatives incorporated into the meal of 4th instar larvae of Rhodnius prolixus. \(\triangle \) Chromane of precocene II/ml of blood; ▲ precocene II/ml of blood; ● ethoxy precocene/ml of blood; ■ precocene II/ml of blood; O precocene II/ml of artificial diet.

level (table 2). Alternative explanations related to suspected biochemical interactions of the precocenes are also possible. The precocenes are presumed to undergo biotransformation into reactive epoxides through the action of monooxygenase enzymes within the corpora allata. These reactive intermediates alkylate cellular elements within the allatal cells resulting in cell death and loss of juvenile hormone secretion 12,24. Many other insect tissues, especially the gut and fat body, contain monooxygenases which may be capable of similar biotransformation of the precocenes²⁵ The antifeeding response induced in Rhodnius following the ingestion of precocene II may be due to a direct cytotoxic action on gut tissues which contain activating monooxygenases. Schooneveld²⁶ has observed degenerative changes occurring within allatal cells within 90 min of precocene treatment. A similar rapidly induced necrosis of gut tissues could explain the diminution in feeding. Since precocene I, II and ethoxy precocene are all potent antijuvenile hormonal agents the difference in antifeeding activity among them might be due to a specificity of the gut monooxygenases for precocene II. The special feeding behavior of²⁷ the hematophagous bug *Rhodnius prolixus* makes it an ideal model to investigate this phenomenon and antifeedant substances in general.

- Acknowledgments. This research was supported by funds from Conselho Nacional de Desenvolvimento Cientifico e Tecnologico, and National Science Foundation grants PCM-76-09647 and PCM-79-03245.
- Reprint requests to W.S.B., Department of Entomology, New York State Agricultural Experiment Station, Cornell Universiy, Geneva (New York 14456, USA).
- W.S. Bowers, in: The juvenile hormones, p.394. Ed. L.I. Gilbert. Plenum Press, New York 1976.
- W.S. Bowers, T. Ohta, J.S. Cleere and P.A. Marsella, Science 193, 542 (1976).
- W.S. Bowers and R. Martinez-Pardo, Science 197, 1369 (1977)
- M.P. Pener, L. Orshan and J. De Wilde, Nature, Lond. 272, 350 (1978).
- G.C. Unnithan, K.K. Nair and W.S. Bowers, J. Insect Physiol. 23, 1081 (1977).
- G. E. Pratt and W. S. Bowers, Nature, Lond. 265, 548 (1977).
- G.C. Unnithan and K.K. Nair, Ent. Soc. Am. 72, 38 (1979).
- H. Schooneveld, Experientia 35, 363 (1979)
- P. Masner, W.S. Bowers, M. Kalin and T. Muhle, Gen. comp. Endocr. 37, 156 (1979).
- W.S. Bowers, in: Insect biology in the future, p.613. Eds M. Locke and D.S. Smith. Academic Press, New York 1980.
- K. Slama, Acta ent. bohemosl. 75, 65 (1978).
- W.S. Bowers and P.A. Ferugia, in: Regulation of insect development and behavior, p.312. Ed. M. Kloza. Wrocław Technical University Press, Wrocław 1981.
- 15 W.S. Bowers and J.R. Aldrich, Experientia 36, 362 (1980).
- W.S. Bowers and D.M. Soderlund, in: Regulation of insect development and behavior, p. 309. Ed. M. Kloza. Wrocław Technical University Press, Wrocław 1981. P.D. Azambuja, E.S. Garcia and J.M.C. Ribeiro, Gen. comp.
- 17 Endocr. 45, 100 (1981).
- W.S. Bowers, Pontif. Acad. Sci., Scripta Varia 41, 129 (1977).
- E.S. Garcia, J.D. Macarini, M.L.M. Garcia and F.B. Ubatuba, An. Acad. brasil. Cienc. 47, 537 (1975)
- 20 G.W. Snedecor, Statistical methods, 5th edn. Iowa State University Press, Ames 1956. W. G. Friend and J. J. B. Smith, A. Rev. Ent. 22, 309 (1977).
- R. F. Chapman, Bull. ent. Res. 64, 339 (1974). 22
- E.A. Bernays and R.F. Chapman, in: Biochemical aspects of plant and animal coevolution. Ed. J.B. Harborne. Academic Press, London 1978.
- G.E. Pratt, R.C. Jennings, A.F. Hamnett and G.T. Brooks, Nature 284, 320 (1980).
- D.M. Soderlund, A. Messeguer and W.S. Bowers, Agric. Food Chem. 28, 724 (1980).
- H. Schooneveld, Experientia 35, 363 (1979).
- P.A. Buxton, Trans. R. ent. Soc. Lond. 78, 227 (1930).